Molecular imaging as the main part of our decision-making and treatment strategies in stroke

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1. ABSTRACT

Review of results of clinical studies indicates the number of potential patients who are actually treated for acute ischemic stroke is disappointingly low, and effective treatments are making a minor impact on this major public health problem. Imaging modalities, such as diffusion- and perfusion-weighted images, as well as CT perfusion and CT angiography, to better select patients for treatment are now routinely performed in most academic medical centers. However, there is not a perfect penumbra imaging technique and each one has its own advantages and disadvantages. Recent advances in molecular imaging modalities allow a better understanding of this pathophysiological process that could lead to enhanced therapy for stroke. This article seeks to describe the role of molecular imaging in identifying the pathophysiology of stroke and how it could be incorporated in future decision-making and treatment strategies in stroke.

2. MOLECULAR IMAGING

Molecular imaging has emerged as a potentially revolutionary discipline that aims to visually characterize normal and pathologic processes at the cellular and molecular levels within the milieu of living organisms. It broadly incorporates methods and concepts from molecular and cell biology, imaging sciences, chemistry, high-throughput biology (e.g., genomics, proteomics), nanotechnology, pharmacology, and bioinformatics (1-3). And through molecular imaging, radiology is expected to play a critical role in advancing molecular medicine and potentially revolutionizing patient care and biomedical research (4, 5).

Conventional clinical imaging generally relies on macroscopic anatomic and physiologic variations for disease diagnosis and assessment. Such morphologic changes are often nonspecific and late phenotypic
manifestations of underlying molecular derangements (1, 6). By contrast, molecular imaging exploits the use of directed imaging probes to sense the specific molecular alterations underlying diseases rather than downstream end effects at the tissue or organ level which means earlier and more precise disease diagnosis, improved disease characterization, and more meaningful monitoring of disease progression.

The impact of molecular imaging in biomedical research is also promising. In vitro research traditionally extracts and studies the events in artificial environments like cell cultures. This is problematic because biologic processes rarely occur in isolation and are instead mediated through a complex and dynamic interplay of gene expression, signaling pathways, environmental factors, and inherent feedback mechanisms. In contrast, biology can be studied under physiological conditions within the intact microenvironment and without any tissue destruction by the application of molecular imaging. This transition from a reductionism to a holistic approach to research (7) enables molecular imaging to be more predictive and relevant in research studies. Molecular imaging also enables real-time monitoring of molecular phenomena through repetitive imaging of an individual subject. Molecular imaging has been rapidly adopted into the basic science research of dynamic biologic processes such as hypoxia (8), inflammation (9), apoptosis (10), angiogenesis (11), tumorigenesis (12), and gene expression (13, 14). More translational applications include stem cell trafficking (15) and monitoring of the distribution and efficacy of novel therapeutic moieties (16). In fact, one of the most promising applications of molecular imaging is to accelerate drug discovery and development.

Imaging modalities used in molecular imaging include positron emission tomography (PET), single photon emission computed tomography (SPECT), magnetic resonance (MR) imaging, optical imaging, and ultrasound (US). These modalities differ in terms of spatial resolution, temporal resolution, sensitivity in probe detection, depth of signal penetration, availability of biocompatible molecular imaging agents, and, of course, cost. Each has its unique advantages and disadvantages, and the choice of imaging system ultimately depends on the question to be addressed (Table 1). These modalities in combination with contrast agents, which are a broad set of molecular probes and tracers that target a specific peptide, receptor ligand, enzyme substrate, oligonucleotide, or antibody, are the basis of molecular imaging technology.

A survey of the molecular imaging literature reveals that its application to human ailments is being explored in a range of disparate fields from neurology and psychiatry to infectious diseases and drug resistance. The goals of this review are to provide a summary of molecular imaging application in stroke and to stimulate discussion among scientists about whether and how it could be incorporated in future decision-making and treatment strategies in stroke.

3. STROKE

Stroke is the leading cause of disability and the third leading cause of death in the USA. Approximately 700,000 strokes occur in the united states annually (17, 18). The clinical burden of stroke and transient ischemic attack now exceeds that of coronary heart disease (19). The American Heart Association (AHA) estimates the cost of treating stroke related injury and disability at $58 billion for 2006 alone (www.americanheart.org).

Treatment of stroke requires a comprehensive understanding of stroke pathophysiology. In ischemic stroke, which represents about 80% of all strokes, decreased or absent circulating blood deprives neurons of necessary substrates. A thrombus or an embolus can occlude a cerebral artery and cause ischemia in the affected vascular territory. The obvious therapeutic response to such a reduction of blood supply is to re-establish flow by pharmacologic or mechanical means as quickly as possible, leading to the advent of the only FDA approved and available therapy for stroke, which is the intravenous (IV) tissue plasminogen activator (tPA) within 3 hours of stroke onset. This treatment is only effective within 3 hours of onset and only treats the symptoms by dissolving the clot. Only 2-3% of stroke patients are able to use this agent because of its narrow therapeutic window. It also has a limited success rate of 35–65% at recanalizing the middle cerebral artery, dropping to about 10% if the internal carotid artery is occluded (20, 21). The disappointing result has been that intravenous tPA for the treatment of stroke has made only a minor impact on a major public health problem. Combined analysis of the tPA clinical trials has demonstrated that the earlier initiation of tPA therapy is associated with improved outcomes which is directly related to the concept of the ischemic penumbra and its evolution (22).

The concept of the ischemic penumbra is familiar to most investigators who work in the stroke field. In brief, the initial definition of the ischemic penumbra by Astrup et al (23), was that portion of the ischemic zone with absent electrical activity but with preserved ion homeostasis and transmembrane electrical potentials. Several revised definitions of the ischemic penumbra appeared over time that focused on thresholds of cerebral blood flow (CBF) decline, energy metabolism and protein synthesis. From the

Table 1. Attributes of molecular imaging modalities

<table>
<thead>
<tr>
<th>Modality</th>
<th>Sensitivity</th>
<th>Spatial resolution</th>
<th>Temporal resolution</th>
<th>Penetration Depth</th>
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<tr>
<td>SPECT</td>
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<td>PET</td>
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<tr>
<td>Optical Imaging</td>
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Adapted with permission from reference 1.
Stroke and molecular imaging

clinical perspective, a relatively simple and straightforward definition of the ischemic penumbra initially proposed by Hakim (24) is that portion of the ischemic region destined for infarction that is currently potentially salvageable with appropriate intervention. This definition provides a framework for approaching the important concepts of evolution of penumbra into infarcted tissue, the time window over which this evolution occurs (i.e. the therapeutic time window) and the use of imaging modalities to identify at least an approximation of this vital ischemic region.

4. NEUROIMAGING AND STROKE

Brain imaging plays a vital role in acute stroke by delineating ischemia from hemorrhage, estimating tissue at risk for infarction and excluding some stroke mimics, such as tumor. Neuroimaging modalities have emerged as integral components of almost all acute stroke trials, which may be used for diagnostic, therapeutic, and prognostic purposes in acute stroke and beyond.

The mainstays of early stroke diagnosis are conventional noncontrast head CT (NCCT) and conventional brain MR imaging (25-33). Emergency, non-contrast CT of the head is the recommended initial neuroimaging study (34). This modality identifies hemorrhage and can help distinguish non-vascular causes of neurological symptoms such as tumor (35). CT is not ideal, however, because of the difficulty in detecting acute or small infarcts and artifact in the brainstem area (34).

In some centers, multiparametric MRI is increasingly used as first-line imaging for patients with suspected ischemic stroke. MRI is better for detection of acute ischemia than CT (36). Diffusion-weighted imaging (DWI) sequences should be done in all acute MRI studies to allow identification of ischemic areas within minutes of symptom onset (37, 38). DWI reveals areas of restricted Brownian motion of water molecules likely due to cytotoxic edema. The sensitivity and specificity of DWI for detecting acute ischemia are about 100%. Diffusion-weighted imaging also provides the additional advantage of visualization of small subcortical lesions and brainstem or cerebellar lesions, usually poorly visualized on CT (39-46). The diagnosis of ischemia may be confirmed by finding corresponding hypointense lesions in a vascular distribution on apparent diffusion coefficient (ADC) maps.

As complementary sequences in a dedicated stroke protocol MRI, magnetic resonance angiography (MRA) and perfusion-weighted imaging (PWI) may provide additional information regarding vessel status, collateral flow, and territories at risk. MRA allows for rapid characterization of the cervical and cephalic large vessels. MRA may delineate extracranial internal carotid artery stenosis with high accuracy compared with digital subtraction angiography (DSA) (47-49).

Dynamic CT perfusion imaging and CT angiography (CTA) have shown remarkable progress in providing information in acute stroke. Dynamic CT perfusion is acquired using sequential imaging in cine mode after intravenous injection of an iodinated contrast medium (50). The data are then processed using either a maximal-slope model or a deconvolution analysis to produce maps of cerebral blood flow (CBF), cerebral blood volume (CBV), time to peak (TTP), and mean transit time (MTT). Dynamic CT perfusion imaging showed good correlation with MRI for CBF and MTT abnormalities in acute stroke (51) and several studies have suggested that CT has the potential of replacing MRI. Nonetheless, DWI remains the sole imaging technique within minutes of onset regardless of location and size, and it is still unlikely to be replaced by CT in the foreseeable future (52). CTA has been demonstrated to be accurate in the evaluation of cervical and large-vessel intracranial occlusion.

Carotid Duplex ultrasound (CDUS) and transcranial Doppler (TCD) ultrasound are noninvasive methods for neurovascular evaluation of the extracranial and intracranial large vessels. Although both methods may help to establish the source of an embolic stroke, they have rarely been used acutely for this purpose. However, accumulating evidence suggests that both Duplex and TCD can be used urgently at the bedside to select patients for interventional thrombolytic or endovascular treatment (53-56).

The penumbra is the target tissue for early thrombolysis and can be visualized by various imaging modalities, including PET, CT and MRI (57-60). PET provides quantitative measurements of CBF, CBV, the metabolic rate of oxygen consumption (CMRO2), and the rate of oxygen extraction (OEF). In PET studies, the penumbra is characterized by the pattern of increased OEF with reduced CBF and CMRO2 identifies penumbral tissue (61). Moreover the PET ligand 18F-fluoromisonidazole (18F-FMISO) is a marker of hypoxic but still viable tissue. Studies with this presumed penumbral marker have confirmed the prolonged existence of penumbral tissue in stroke models and patients (62). In MRI studies, the equivalent of the penumbra is called “mismatch”, which can be visualized by a combination of DWI and PWI (63). Minutes after the onset of stroke, MR DWI can demonstrate restriction of the random motion of water and decline of its ADC (64). This is being used as a marker of irreversible neuronal damage resulting from critically severe tissue hypoperfusion. Moreover, identifying a mismatch between such a diffusion lesion and the extent of the perfusion abnormality on PWI has in turn evolved as a means of depicting the potentially viable penumbra. The CBF/CBV mismatch on perfusion CT correlates well with regions of DWI/PWI mismatch when both studies are obtained in a close temporal window (65). The validation of the thresholds required to identify the abnormal CBF and CBV regions remains to be proven. Other problems with perfusion CT include the need to inject iodinated contrast to obtain the images and the relatively restricted tissue volume that can be imaged currently. DWI/PWI MRI can essentially image the whole brain and is not restricted to several slices as perfusion CT currently is. Another problem with perfusion CT as compared to DWI/PWI is the tracking of presumed infarct volumes over time.
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Figure 1. Comparison of horizontal T2-weighted image (A), isotropic diffusion-weighted image (B), absolute magnetization transfer ratio asymmetry image (C), absolute pH image (D), and 2,3,5-TTC-stained histology (E) of an ischemic rat brain. The ischemic area (arrow) is located in the caudate nucleus, a region that commonly becomes affected after occlusion of the MCA. No effect of ischemia was visible on the T2-weighted image (A), but the pH-sensitive images (C and D) show the ischemic region, as confirmed by the diffusion-weighted image (B), and by histology, which was acquired 8 h later (E). Adapted with permission from ref. 72.

Although currently diffusion and perfusion MRI is widely used in clinical trials to approximate the location and persistence of the ischemic penumbra, to select appropriate patients for inclusion and to evaluate the treatment effect (52), recent evidences which have route in molecular imaging are questioning many of its principal assumptions.

Lin et al. (66) explored the relationship between CBF and ADC and showed that after an initial gradual decline, an abrupt and significant reduction of ADC occurs at CBF values around 15 mL/100 g/min. However, the threshold was time dependent, rising to 24 mL/100 g/min at later time points. Beyond method-related errors, this time dependence may explain why some studies have reported ADC changes at CBF as high as 50 mL/100 g/min and explain the difficulty of identifying an absolute ADC threshold for irreversible damage (67). On the other hand, Nicoli et al. (68) using MR spectroscopic imaging demonstrated that within regions of severe ADC decrease, a heterogeneous pattern of metabolic injury exists, evidenced by wide variations in lactate and N-acetyl aspartate ratios. Also, in a recent study using multitracer PET, Guadagno et al. (69) documented that the DWI lesion does not represent a single stage of the ischemic process and that it incorporates potentially salvageable penumbral tissue. They demonstrated a heterogeneous pattern of blood flow, oxygen metabolism, and extraction fraction within the DWI lesion, inconsistent with the notion of uniformly irreversible neuronal damage. Furthermore, in a subsequent report, they showed that severe ADC reduction existed in the penumbra as well as the core (70), and thus cannot solely represent irreversible damage. Loh et al. (71) have also found areas of reduced ADC outside of the acute DWI lesion that did not progress to infarction and thus argue against an absolute threshold based on ADC alone.

Recently, Zhou et al. (72) developed a new MRI approach based on a magnetization transfer (MT) experiment in which the exchangeable amide protons of mobile tissue proteins and peptides are selectively irradiated using radiofrequency. This causes saturation of these protons, which is subsequently transferred to the water protons. Even though the concentration of such cellular proteins and peptides is in the micro- to millimolar range, these effects can be detected as a few percent signal change in the water line through a sensitivity enhancement mechanism. Sun et al. (73) used this technique to acquire pH-weighted images during acute ischemia and showed that the outer boundary of the hypoperfused area that shows a decrease in pH without DWI abnormality may better correspond to the classic ischemic penumbra border than the PWI deficit (Figures 1 and 2).

In a recent pilot study, Blankenberg et al. using 99mTc-HYNIC-annexin V SPECT imaging in acute stroke showed that human image data of that annexin V uptake correlates to sites of restricted diffusion on MRI. The regions of restricted diffusion seen on MRI also appeared to be more uniform than the heterogeneous multifocal uptake of annexin V. This may imply that annexin V imaging better reflects sites of varying degrees of brain injury as compared with DWI MR imaging on a molecular level (74). Thus, the current uncertainties surrounding MR diffusion-perfusion imaging indicate that the “mismatch” does not always “match” the penumbra (58, 66-71, 73-75). The key question is “how important is the molecular imaging role in penumbra imaging and stroke treatment?”

4.1. Molecular imaging of the penumbra

A better identification of the true penumbral tissue might rely on the cascade of molecular events started after vascular occlusion that have been the objective of extensive research in recent years and development of new strategies for stroke treatment hinges on better understanding of this complex cellular and molecular interplay that ensue following stroke. Thus, penumbra as the primary target of acute stroke therapy suggested that imaging identification of the penumbra could be of great value in the selection of patients for acute stroke therapy.

4.1.1. Molecular imaging of hypoxia

Recently, 18F-FMISO PET imaging of stroke patients has been investigated as a possible penumbral imaging method, which, because of the direct imaging of cellular hypoxia, may overcome some of the difficulties with existing indirect perfusion-based imaging methods. Nitroimidazole compounds such as 18F-FMISO diffuse freely across cell membranes and in living cells are then reduced by intracellular reductases into a radical anion. In normoxic conditions, this compound is rapidly reoxidized and diffuses back out of cells. Under hypoxic conditions, that is, inadequate oxygen supply relative to demand, however, further reduction steps occur, and the reduced compound becomes irreversibly bound to intracellular macromolecules. Thus, reduced nitroimidazoles are supposed to be trapped within hypoxic cells, but not by necrotic or nonhypoxic cells (76).

Human 18F-FMISO studies have been revealing about the temporal and spatial evolution of hypoxia in
stroke and in correlating these changes with clinical outcome. However, there are some unresolved questions that are not easily answered in human studies. As, is binding specific to hypoxic tissue, or will there be ongoing binding after reperfusion (either spontaneous or after thrombolytic therapy)?

Takasawa et al. (77) in a recent pilot study in rats, showed that trapping of $^{18}$F-FMISO in the stroke area only occurred in the early phase of middle cerebral artery occlusion (MCAO), but not when tissue necrosis has developed (i.e. 48 h after permanent MCAO) or if early reperfusion has occurred, which strongly support the validity of $^{18}$F-FMISO as a specific marker of the viable hypoxic brain/penumbra after stroke (Figure 3).

4.1.2. Molecular imaging and energy metabolites

The concentration of energy metabolites including ATP, phosphocreatine, lactate and $n$-acetyl aspartate (NAA), differs between the ischemic core and the penumbral tissue where the metabolic concentration decrease is less severe (78) so the use of a molecular imaging technique might be helpful to characterize the ischemic penumbra in individual patients.

MR spectroscopy (MRS) has been used in vivo to estimate the concentration of certain metabolites such as creatine, choline, NAA, lactate, myoinositol, glutamate/glutamine, and lipids (79). The physical basis of MRS is the chemical shift effect, that is, the fact that nuclei located in different molecular environments (different molecules or different locations within a molecule) sense slightly different magnitudes of magnetic field, causing them to precess at different rates.

MRS provides plots (spectra) of signal intensity, which is proportional to concentration, versus precession rate shift with respect to a reference, expressed in parts per million (ppm). Even though any nucleus with nonzero spin could in principle be studied, clinical MRS practice is most commonly based on the hydrogen nucleus. Molecular groups generate specific resonance patterns on the spectrum, either as single peaks, doublets, or more complex shapes. A given molecule therefore may have multiple corresponding peaks, only some of which may be observable. As magnetic field strength is increased, the separation of peaks improves, increasing the number of metabolites that can be distinguished. Especially at low field strengths, observed peaks may contain contributions...
Figure 3. Positron emission tomography data and histology in the permanent MCAo rat. (A) Early phase (sum of first 10 frames, that is, 0 to 5 mins) and late phase (sum of last 10 frames, that is, 130 to 180 mins) for the first PET session (started 39 mins after clip on), showing prominent late trapping of the tracer in the affected cortex (arrow), and some trapping as well in the surgically injured temporalis muscle (arrowheads); (B) Time–activity curves (TACs) for the affected and unaffected sides for the first PET session. (C) Early phase and late phase accumulated images for the second PET session (48 h later; same frames as first session), showing symmetrical late retention; (D) TACs for the affected and unaffected sides for the second PET session. (E) Fusion images with MRI template (coronal images), also illustrating the affected cortex ROI on two slices (see Materials and Methods), and the surgically injured temporalis muscle (arrowheads); (F) illustrative Cresyl Violet (CV) coronal sections at approximately the same levels as in (E); (G) CV at high magnification. Adapted with permission from ref. 77.
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from several affine molecular types and locations. MRS studies can be carried out for a single voxel or for a matrix of voxels. Due to a number of confounding measurement variables, reliable quantification is very difficult to achieve; therefore, concentrations are usually expressed in arbitrary units, or as ratios. As the chemical shift in ppm is constant for all peaks, prior knowledge can be used for their identification and fitting.

A parameter known as echo time (TE) controls the delay from the radiofrequency excitation pulse to the detection of the echo. Due to different T2 relaxation times among metabolites, changing the TE changes the appearance of the spectrum. Short echo times (on the order of 20-40 ms) result in spectra containing contributions from a large set of metabolites and macromolecules, which may be difficult to separate; long echo times (on the order of 120-300 ms) result in spectra with a flatter baseline, on which only creatine, choline, NAA, lactate, and lipids are visible (80).

Detection of lactate by in vivo proton MRS may provide useful information on metabolic stress in brain, potentially identifying the degree of ischemia (81-84). Therefore, if lactate can be quantitatively evaluated, it could have an important clinical impact. However, a large part of the lactate signal overlapped with lipid signals in conventional MRS measurements (85, 86). Thus, lactate has been thought to rise early after the insult in the acute phase (<24 h) and may remain high over a long period into the chronic phase (>7 days) (87), which could lead us to an inappropriate understanding of the pathophysiological status in the ischemic lesion.

Harada et al. (88) in a recent study using 7 Tesla MRS and stimulated echo acquisition mode (STEAM) with longer TE demonstrated that MRS-detectable lactate increased rapidly in the infarcted rat brain 6 h after MCAO and disappeared within 48 h following MCAO. In addition, MRS detectable lipid was also observed 6 h after MCAO and continued increasing over 1 week, which was confirmed by histological examination.

Thus, MRS at higher field strengths provides better signal to noise ratio (SNR) and increased spectral, spatial and temporal resolution, allowing the acquisition of high quality, easily quantifiable spectra in acceptable scan times (88). This may provide precise biochemical information from distinct regions of the brain noninvasively that is clinically useful by evaluating the severity of cerebral ischemia and the status of the patient in response to the therapy.

Moffet et al. (89) have recently summarized evidence from numerous studies showing that reduced NAA levels as detected by MRS can be an extremely valuable marker of brain injury after stroke or hypoxia. In vivo MRS studies support the hypothesis of NAA as a surrogate for neuronal loss and dysfunction, and the clinically associated neurological deficits observed in patients after local or global hypoxia-ischemic incidents. However, several key issues including the basis of the relatively slow observed decreases in NAA signal after the onset of stroke and whether NAA levels can recover over time under certain circumstances remain unresolved.

4.1.3. Molecular imaging of glutamate-mediated channels

The decrease in the CBF leads to a marked reduction in ATP with Na+/K+ pump failure and an increase in extracellular glutamate which activates glutamate-mediated channels and results in neuronal damage and eventual death.

Glutamate stimulation of α-amino-3-hydroxy-5-methyl-4-isoxazolone-proprionic acid (AMPA) receptors increases sodium uptake by neurons and cellular edema (90). However, glutamatergic stimulation of N-methyl-D-aspartate (NMDA) receptors by glutamate increases intracellular calcium concentrations and cell death (91, 92). NMDA receptors exist as tetrameric subunit assemblies made from both NR1 and NR2 receptor subunits (93-96). The NR2 subunits (NR2A-D) give rise to four receptor subtypes, which have different pharmacological properties and expression patterns in the CNS (97). NR2A and NR2B are the predominant NR2 subunits in the adult forebrain, where stroke most frequently occurs. These subunits have differential roles in supporting neuronal survival and mediating neuronal death and hence have opposing impacts on excitotoxic brain damage after acute brain insults such as stroke and brain trauma. Recently several studies have strongly suggested that it is the NR1/NR2B, not the NR2A-containing, NMDA receptor subpopulation that plays a primary role in triggering intracellular cascades that lead to NMDA- or ischemia-induced neuronal apoptosis and in fact, NR2A is involved in the NMDA-mediated neuronal survival (98-100). Such opposing actions may explain, at least in part, the failure of NMDA receptor antagonism-based clinical trials of stroke and provide a scientific basis for developing novel and effective NMDA receptor-based stroke therapies.

PET tracers targeting the NR2B receptor, by allowing non-invasive imaging in the living brain in stroke models, could facilitate drug discovery and provide new diagnostic tools. Over the last few years several compounds have been synthesized and evaluated as NR2B-selective PET tracers. Despite promising results obtained in vitro, all compounds evaluated so far have suffered from low to moderate brain penetration and lack of NR2B-specific binding in vivo. Recently, Merck published a new series of NR2B-selective compounds, based on the N-benzyl amidine structure and Arstad et al. (97) in a study using three different compounds of N-benzyl amidine class, demonstrated 11C-labelled N-benzyl amidines have a brain uptake suitable for PET imaging. In addition, they were also found to have favorable uptake and retention in the brain, as well as a binding pattern consistent with the expression of the target receptor as measured by in vitro autoradiography. However, the metabolism of the compounds tested was too rapid to allow for in vivo imaging. Further optimization of this class of compounds may lead to successful PET tracers for the NMDA NR2B subtype receptor (97).
4.1.4. Inflammation and molecular imaging

Local brain inflammation is a pathologic hallmark of ischemic stroke lesions (101, 102) and is spatiotemporally related to the occurrence of delayed apoptotic cell death (103). Phagocyte cells, involving resident microglia and infiltrating macrophages, have both protective and destructive role in stroke. Microglia is activated within minutes of ischemia onset and produces a plethora of inflammatory mediators, which exacerbate tissue damage (104-106), but may also protect the brain against ischemic and excitotoxic injury (107-109). In contrast to the rapid microglia response, blood-derived macrophages are recruited with a delay of at least 24 to 48 hours (110-112). At these later stages, the synthesis of proinflammatory cytokines is already down regulated (113), whereas various anti-inflammatory and protective factors are progressively expressed (114, 115). Thus, inflammatory responses to -brain ischemia are heterogeneous and at cellular level, phagocyte cells represent a potential therapeutic target. To date, most studies on microglia/macrophage response to cerebral ischemia have used cell culture or ex vivo histology and several studies in experimental stroke models showed that multimodal MRI comprising diffusion- and perfusion-weighted as well as gadolinium-enhanced imaging is not able to discriminate inflamed from noninflamed infarct subareas (116). Thus, new molecular imaging techniques to monitor phagocytic activity in brain ischemia need to be developed.

Already several years ago, superparamagnetic iron oxide nanoparticles have been introduced as cell-specific contrast agents that can be injected intravenously and are taken up by cells of the mononuclear phagocyte system (117). Based on particle size, one can distinguish superparamagnetic iron oxide (SPIO; \( \approx 60 \) to 150 nm diameter) from ultrasmall superparamagnetic iron oxide particles (USPIO; \( \approx 20 \) to 50 nm). SPIO particles are rapidly phagocytosed by cells of the reticuloendothelial system in liver and spleen leading to rapid clearance from the blood pool after intravenous injection. In contrast, circulation times of USPIO are considerably longer which, at least in theory, favors the interaction with phagocyte populations in the blood and lymph nodes. USPIO nanoparticles cause microscopic field gradients that efficiently diphasic nearby protons by disrupting the homogeneity of the magnetic field, thereby strongly enhancing the transverse relaxation times T2 and T2* over a length scale much larger than the nanoparticles’ size. Hence, they are called T2 contrast agents, and they decrease signal intensity in standard imaging sequences (117). Their relaxivity increases with field strength up to its saturation threshold, with higher MR field strengths producing larger signal intensity changes and hence a higher contrast-to-noise ratio. Accordingly, USPIO agents such as Ferumoxtran-10 (Combidx, Sinerem) are in advanced clinical development for lymph node imaging (118). whereas SPIO particles such as ferumoxide (Endorem, Resovist) have been approved for liver imaging. Despite these differences, both types of contrast agents have been successfully used in the study of inflammatory processes in brain ischemia. (119-124)

In experimental brain ischemia, Rausch et al. (119) were the first to use USPIO for macrophage imaging in a model of permanent middle cerebral artery occlusion (pMCAO). In this study, ferumoxtran-10 was injected intravenously into rats at 5 hours after pMCAO followed by repeated MRI on days 1, 2, 4, and 7. On T2-weighted images, patchy areas of signal loss in the infarctions were found until day 4 and decreased thereafter. Essentially similar findings were obtained in a transient ischemia model (120). Using photochemically induced ischemia model, several studies showed accumulation of iron oxide particles in the infarct border zone during subacute stages of lesion development (121, 123, 124). Importantly, iron-related signal changes on MRI were paralleled by macrophage-associated iron deposition detected histochemically on postmortem brain sections. In the study by Kleinschmidt et al. (121), SPIO injection between days 5 and 6 after ischemia, but not at earlier time points, caused typical signal loss on T2*-weighted images obtained 24 hours after injection. Thus, based on the assumption that SPIO was primarily taken up by circulating phagocytes, this study suggests that infiltration of SPIO-laden macrophages occurred to a significant extent only at the end of the first week after ischemia. This is in line with earlier results suggesting that the recruitment of hematogenous macrophages likewise occurs in a narrow time interval between days 3 and 6 after ischemia (110-112). Collectively, these findings demonstrate that appropriate timing of contrast agent injection and subsequent MRI is of critical importance for iron oxide particle-based macrophage imaging in brain ischemia.

Microglia and macrophage responses are segregated not only with respect to time, but to some extent also spatially. After focal cortical ischemia, there is secondary involvement of the ipsilateral thalamus attributable to retrograde degeneration of thalamocortical projection fibers (125-127). In these areas of delayed degeneration, strong and long-lasting microglia activation develops, whereas hematogenous macrophages are largely excluded. In the studies reported so far, iron oxide particle-related signal changes were restricted to the primary lesion site and not observed in the thalamus. These observations further support the concept that particle uptake occurs peripherally with subsequent infiltration of iron-laden cells into the lesioned central nervous system parenchyma. Thereby, iron oxide particle-enhanced MRI potentially differs from PET with \(^{11} \text{C-PK11195}\) as a radioactive ligand to peripheral-type benzodiazepine-binding sites on mononuclear phagocytes. Previous studies showed increased binding of \(^{11} \text{C-PK11195}\) in the thalamus ipsilateral to a cortical MCA infarction indicating that \(^{11} \text{C-PK11195}\) PET detects microglia responses in the degenerating areas (126, 127). USPIO uptake and \(^{11} \text{C-PK11195}\) binding may therefore reflect distinct aspects of the cellular inflammatory response to brain ischemia.

Recently Jander et al. (122) conducted a trial of USPIO-enhanced MRI in 10 consecutive patients with early ischemic stroke. In their study, USPIO contrast agent was infused 5 to 6 days after stroke onset, which corresponds to the presumed period of hematogenous macrophage recruitment (110, 121) and infarct expansion shown in
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quantitative MRI analysis (128, 129). In line with preclinical safety studies (130), USPIO infusions were well tolerated by all patients. To delineate lesion extension, all patients underwent initial stroke MRI, including diffusion- and perfusion-weighted imaging, within 24 hours after stroke. To define the pattern of gadolinium-based contrast medium enhancement, a first follow-up gadolinium-enhanced MRI was performed 4 to 5 days after symptom onset (i.e., 24 hours before USPIO infusion). Two additional scans were obtained at 24 and 48 hours after USPIO infusion. As the main finding, they observed consistent USPIO-related signal changes in all 10 patients, whereas gadolinium enhancement only occurred in three patients. Thus, USPIO enhancement was not a mere epiphphenomenon of blood–brain barrier breakdown. Interestingly, their study revealed two distinct components of USPIO-related signal changes, one associated with blood vessels and one representing parenchymal enhancement. Vessel-associated changes appeared as signal loss on T2/T2*-weighted images and decreased from the first to second scan after USPIO infusion, most likely reflecting a transient blood pool effect of the contrast agent. Conversely, parenchymal enhancement was mainly evident on T1-weighted images, increased over time, and matched with the expected distribution of macrophages. They therefore suggest that the increasing USPIO enhancement on T1-weighted images indicates brain infiltration by USPIO-laden macrophages. Similar to these findings, Dousset and colleagues (131) reported prominent signal changes on T1- rather than on T2*-weighted images in patients with multiple sclerosis. The reasons for the distinct USPIO-related signal patterns on T1- versus T2*-weighted images are currently unknown.

These studies raise the perspective that USPIO-enhanced MRI may provide a novel in vivo surrogate marker of cellular inflammation in stroke and other central nervous system pathologies. There are a number of open questions that require further study. First of all, the assumption that circulating monocyte-derived phagocytes act as the principal cell type responsible for iron particle uptake after IV injection is so far only based on indirect evidence. Furthermore, even circulating mononuclear phagocytes can be subdivided into subpopulations with distinct phenotypic and functional profiles (132). Thus, USPIO enhancement may reflect the infiltration of a specific macrophage subpopulation, which remains to be identified. Finally, inflammation may not only have deleterious consequences for ischemic lesion progression, but may also mediate beneficial effects such as lesion demarcation, wound healing, and tissue regeneration. So far, the specific functional contribution of macrophages for the development of brain infarction is essentially unknown. USPIO-enhanced MRI may represent an important tool to address this issue both in basic science and clinical studies.

An alternative strategy to image inflammation is to develop MRI contrast agents specifically targeting an inflammatory cellular marker like the endothelial adhesion molecules. E-selectin (CD62E, ELAM-1) is an adhesion molecule expressed on the luminal surface of vascular endothelial cells in inflammation. It serves the function of initiating interactions between the endothelial cell and circulating leukocytes, preceding leukocyte diapedesis into inflamed tissue (133).

Chapman and colleagues showed that anti–E-selectin monoclonal antibodies (MoAbs) labeled with indium-111 or technetium-99m can be used to image activated endothelium in vivo in porcine monoarthritis (134-136), as well as inflamed tissues in rheumatoid arthritis (137-139) and inflammatory bowel disease (140). Importantly, radiolabeled anti–E-selectin imaging was much more sensitive than imaging with radiolabeled nonspecific immunoglobulin, demonstrating the advantage of this approach to investigating inflammation over methods that rely on increased endothelial permeability (137).

Exposure to ionizing radiation and the relatively poor anatomic resolution of nuclear gamma cameras limit the value of radiolabeled antibodies in imaging. Therefore, other modalities, including MR imaging (141, 142) and near-infrared optical imaging (143), have been assessed in vitro for E-selectin targeting. Furthermore, sialyl Lewis X, a carbohydrate moiety that binds E-selectin, has been conjugated to gadolinium and used to target E-selectin at MR imaging in models of focal brain ischemia (144) and cytokine-mediated inflammation (145). As USPIO nanoparticles degradation occurs through normal physiologic iron-handling pathways (146), it has marked potential safety advantages over gadolinium, which has no known intracellular excretion pathway. Kang et al. (141) modified USPIO nanoparticles to bind E-selectin in vitro and recently Reynolds et al. (147) used an antibody for targeting USPIO, they used F(ab’2)2 fragments to minimize interactions with Fe receptors or complement. Antibody conjugation of USPIO provides proof-of-principle for the efficacy of targeting E-selectin for in vivo MR imaging, allowing a comparison in due course with other, possibly less specific, ligands that bind E-selectin, such as peptides (148, 149) or carbohydrates (144, 145). They showed that antibody-conjugated USPIO can be used for MR imaging of E-selectin expression on vascular endothelium in vivo. This could be valuable for diagnosing and monitoring early or occult inflammation and may provide an attractive alternative to established investigations such as those involving radiolabeled leukocytes (150).

4.1.5. Molecular imaging of apoptosis in stroke

Apoptosis, or programmed cell death, has also been suggested as being a contributor to the evolution of the penumbra to necrotic tissue. As protein synthesis is required for apoptosis to occur, it is likely that this mechanism only contributes to the progression of the ischemic lesion in areas of modest ischemia or after reperfusion (151). Therefore, molecular imaging of apoptosis can be a highly useful tool in monitoring of stroke course and its response to treatment.
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Imaging of apoptosis in stroke has been attempted with radio-labeled annexin V, a 36 kDa protein that binds to phosphatidylserine (PS), exposed on the cell surface of the apoptotic cell. However, annexin V in stroke imaging was found to underscore the infarct region, while showing multiple bilateral foci of increased signal, extending beyond the regions damaged by the ischemic insult (74, 152, 153). This questionable selectivity and specificity of annexin V for imaging of cell death in stroke may be attributed, at least in part, to its being a relatively large protein, with limited brain access and slow blood clearance. In addition, this probe binds in early apoptosis only on the cell surface, without intracellular uptake and accumulation (154), thus potentially reducing signal/noise ratio.

ApoSense is a novel family of low-molecular weight compounds, designed to address the challenge of apoptosis imaging in vivo. It comprises amphipathic compounds that do not cross the plasma membrane of intact viable cells, but perform selective passage through the membrane of apoptotic cells and accumulation in the cytoplasm from the early cellular stages of the death process. Few studies have shown imaging of cell death, both in vitro and in vivo, by two members of the ApoSense family: DDC (N,N' didansyl-1-cystine, MW = 707) (155), and NST-732 [(5-dimethylamino)-1-naphthalene-sulfonyl-n-ethyl-fluoroalanine, MW = 368] (156). Upon intravenous systemic administration in vivo in animal models of disease-related apoptosis, these compounds have been shown to target specifically the cells undergoing cell death, with marked intracellular accumulation.

Reshef et al. (157) recently used DDC as a probe for detection of cell death in experimental stroke in vivo. DDC manifested selective uptake only by brain areas in the territory of vascular compromise (MCA in this case), and not by other parts of the brain. Within the affected region, uptake was selective only into cells undergoing cell death, with intracellular accumulation of the compound. A sharp border was outlined between intact and cell death regions of the brain. Interestingly, within the same region, conceivably sustaining similar ischemic insult, a mosaic of cells undergoing cell death and intact cells was found, which is characteristic of apoptosis. Detection of cell death by DDC in this study was similar to the pattern of DDC uptake observed in other models of cell death in vivo, such as renal injury (155). This study showed the capability of DDC to delineate differential load of cell death in various regions of the infarct. This analysis of the infarct region, according to the differential load of cell death in its various regions, can be useful for accurate and sensitive assessment of the extent of tissue damage and potential reserves of salvageable tissue within the areas suffering from the vascular compromise.

4.1.6. Angiogenesis and molecular imaging

Reperfusion and collateral revascularization of potentially viable tissue is an important factor in determining patient recovery. Angiogenesis within the penumbra begins after the onset of ischemic stroke and the extent of angiogenesis is correlated with a patient’s survival time (158). Although angiogenesis is stimulated by hypoxia, but how the entire process affects outcome after focal cerebral ischemia is still unclear. It has been proposed that angiogenesis resulting from local hypoxia inhibits the degradation of hypoxia-inducible factor-1 (HIF-1), allowing its sustained expression and stimulating expression of vascular endothelial growth factor (VEGF).

Angiogenic growth factors are secreted by infiltrating macrophages, leucocytes and damaged blood platelets, and this probably helps to maintain increased circulatory expression after stroke. Increased VEGF expression was reported in human brain tissue following acute ischemic stroke (159), whilst Issa et al. (160) suggested that bFGF (FGF2) upregulation is one of the mechanisms that leads to angiogenesis and neuroprotection in the penumbra region after acute stroke. VEGF, one of the most potent angiogenic factors, is upregulated within hours of stroke and has a strong influence on the growth of new blood vessels after ischemia. Certainly, angiogenesis offers a most promising target for both prevention and amelioration of the devastating consequences of acute ischemic stroke in humans. Furthermore, new studies have suggested that injured vessels, activated vessels, and new vessels produce an array of trophic factors and cytokines, including vascular endothelial growth factor and brain-derived neurotrophic factor, which evoke neurogenesis (161-164) are chemotactic for progenitor cells, and enhance the survival and integration of neuroblasts into the cerebral tissue. These factors also facilitate synaptic activity and plasticity (161) and may have major applications in future cell therapies for stroke.

Although increased expression of growth factors and cytokines occurs in many stroke models, many of the signaling pathways that are responsible for revascularization and neurogenesis have only been subjectively proposed on the basis of in vitro studies. Thus, molecular imaging may have a pivotal role in better understanding of these pathophysiological pathways.

VEGF and integrins (particularly αvβ3 integrin) have been identified as favorable targets for imaging angiogenesis, we and others have focused on the development of probes for these specific targets. Although, multimodality imaging of VEGF expression has been reported for many diseases such as cancer, myocardial infarction and ischemia, but in vivo imaging of VEGF/VEGFR expression in stroke has been surprisingly understudied.

4.2. Molecular imaging in stroke prevention

Effective stroke prevention is vital to reduce its clinical burden. Accurate carotid imaging is important for effective secondary stroke prevention. There is a need for new adjuncts to risk stratify patients with carotid artery disease as symptomatology and the degree of luminal narrowing, which have been previously used as clinical grounds for surgical intervention, may no longer be appropriate to be solely considered for decision making. Part of the reason for this is that luminal stenosis alone has
The composition and stage of atherosclerotic plaques are clinically more relevant than the severity of stenosis to evaluate the risk of acute ischemic events. The vulnerable carotid atheromatous plaque, considered responsible for many acute ischemic events, usually has a thin fibrous cap, a large lipid pool and macrophage-dense inflammation on or beneath its surface (166). In contrast, stable or "safe" plaque is fibrous, with little lipid and little or no inflammation. Inflammation within atherosclerotic lesions is likely to increase the risk of plaque rupture and subsequent thrombo-embolism (167) and therefore is an obvious target for novel plaque stabilization interventions. Hence there is the need for the development of new imaging techniques for the detection of vulnerable plaques and particularly plaque inflammation.

It has been possible to image carotid plaque inflammation with USPIO enhanced MRI in both animal (168) and in vivo human studies (169). The use of a USPIO agent – Sinerem™ (Guerbet, Roissy, France), has allowed the direct visualization of macrophage infiltration of carotid atheroma in vivo (169-172). USPIO particles are taken up by activated macrophages in vulnerable plaques and, when clumped in the phagolysosomes produce a strong T2* susceptibility effect, visible on T2* weighted sequences as magnetic susceptibility artifacts (MSA) or signal voids.

In addition to the T2* effect, there is a predominant T1 shortening effect at low concentrations of USPIO in circulating blood, allowing better visualization of the fibrous cap and suggesting that this contrast agent could be used to detect not only inflammation within vulnerable plaque but also aid in identification of "safer" plaque with a significant fibrous component (171). There are also several studies that have used 18F-fluorodeoxyglucose (18F-FDG) PET imaging for noninvasive measurement of atherosclerotic plaque inflammation (173, 174) (Figure 4).

Biomechanical stress is also considered to be a major determinant of plaque vulnerability (175). Structural analysis has suggested that triggers of plaque rupture include high von Mises stress concentrations on a thin fibrous cap and low vessel wall shear stresses as a result of turbulent blood flow (176, 177). Although physiologically important in a purely biomechanical study, wall shear stress is relatively minor compared with the internal stresses of the plaque (von Mises stresses). Finite element analysis (FEA) techniques has been used successfully to perform stress analysis of vulnerable carotid plaques based on the geometry derived from in vivo MRI (178). Li et al. (179) showed that the maximal stresses in the symptomatic plaques were higher than those in the asymptomatic ones and they consistently found that the peak stresses within the shoulder regions and thinnest areas of fibrous cap, of the plaque, which seems to support the notion that the location of plaque rupture is associated with the location of the peak stress. Likewise, the authors showed that, in an idealized model of the carotid bifurcation, it is fibrous cap thickness rather than overall luminal stenosis that appears to contribute most to the overall plaque stress.

Recently Tang et al. (180) performed a study to investigate whether there is a relationship between the degree of MR-defined inflammation using USPIO and biomechanical stress using FEA, in carotid atheromatous plaques. They have demonstrated that there is a significant relationship between biomechanical stress and USPIO
USPIO and macrophage number remains unclear. The relationship between the extent of signal drop seen post-USPIO uptake in inflammatory carotid atheroma and the consensus in the literature as to how best to quantify plaque inflammation and plaque stress is a linear one. This may be difficult to elucidate when there is currently no plaque inflammation and 1 week to 6 weeks indicate weeks after transplantation from a representative rat. Prussian blue staining was used to identify superparamagnetic particle labeled cells on coronal sections. This staining reacts with iron to produce blue color. The Prussian blue staining sections (C) obtained from the same rat 6 weeks after transplantation showed clusters of Prussian blue positive cells indicating superparamagnetic particle labeled cells (see C, black arrows) in the ischemic boundary. These blue cells were localized to angiogenesis areas (C, white arrows). Panels D and E are the vWF immunoreactive images of coronal sections, which matched MRI sections from the same animal sacrificed at 6 weeks, showed an increase in numbers of vWF immunoreactive vessels (left image in D, black line area; left image in the magnified vWF immunostained image E, arrows) in vWF immunostained images. Adapted with permission from ref. 197.

Figure 5. Dynamic migration of transplanted cells in ischemic brain: The dark areas in the 3D MRI were not detected before transplantation of labeled progenitor cells (column Before in A to B). In contrast, the same rat exhibited MRI signals at the cistern at 1 day after injection of superparamagnetic particle labeled cells (A, 1 day, arrow). The dark areas in the 3D MRI first reached the ipsilateral striatum nearby the ipsilateral lateral ventricle within 1 week after transplantation (B, 1 week, arrow). The dark areas in the ipsilateral striatum were clearly visible during the 6 week period. Dark areas were not detected in the contralateral hemisphere at any time points after transplantation (B, 1 day to 6 weeks). Panels A and B represent different levels of coronal sections from the posterior to anterior brain (A, bregma_1.3 mm; B, bregma_1.3 mm). Before and 1 day represent 1 day prior to and 1 day after cell transplantation, and 1 week to 6 weeks indicate weeks after transplantation from a representative rat. Prussian blue staining was used to identify superparamagnetic particle labeled cells on coronal sections. This staining reacts with iron to produce blue color. The Prussian blue staining sections (C) obtained from the same rat 6 weeks after transplantation showed clusters of Prussian blue positive cells indicating superparamagnetic particle labeled cells (see C, black arrows) in the ischemic boundary. These blue cells were localized to angiogenesis areas (C, white arrows). Panels D and E are the vWF immunoreactive images of coronal sections, which matched MRI sections from the same animal sacrificed at 6 weeks, showed an increase in numbers of vWF immunoreactive vessels (left image in D, black line area; left image in the magnified vWF immunostained image E, arrows) in vWF immunostained images. Adapted with permission from ref. 197.

enhanced MR-defined inflammation within carotid atheroma. While there is an obvious association between USPIO and predicted plaque stress from FEA, this of course does not necessarily imply a causative relationship. It remains unclear whether or not the correlation between plaque inflammation and plaque stress is a linear one. This may be difficult to elucidate when there is currently no consensus in the literature as to how best to quantify USPIO uptake in inflammatory carotid atheroma and the relationship between the extent of signal drop seen post-USPIO and macrophage number remains unclear.

Complex biological systems such as atheromatous plaques need to be considered in terms of both their physical mechanical properties and their pathophysiology. The complex interplay between cellular biology and biomechanics is emerging as an extremely important concept for the further understanding of pathological processes and MR imaging will continue to play a major role in uncovering these poorly understood relationships.

4.3. Stem Cell and Stroke Therapy: The role of Molecular Imaging

Stem cells have been used to treat neurological diseases in which neuronal death is the major pathogenetic mechanism, including cerebrovascular and other neurodegenerative diseases (181). Moreover, the first human neural stem cell clinical trial has recently been approved for Batten disease, a pediatric lysosomal storage disease that leads to neuronal loss and death (182). With such advances it becomes important to answer critical questions about the prospects of cell transplantation for stroke therapy.

Stem cells have the capacity to self-renew and differentiate into different cell types, including neurons, astrocytes, and endothelial cells. Stem and progenitor cells are present in fetal cells, immortalized cell lines, umbilical cord blood, bone marrow, and specific organs, including the brain. Animal studies suggest that stem cells (including those from bone marrow) can survive, integrate, and function as neurons in experimental models of stroke (183-186). Because the application of stem cell-based therapies is potentially wide-ranging, specific methods are needed to continuously and noninvasively monitor stem cell survival.

Three-dimensional imaging and in vivo cell tracking capabilities allow MRI to provide high-resolution visualization of the fate of cells after transplantation, and the migration of cells after injection (187) (Figure 5). Recently Shin et al. used Gd-DTPA as MRI contrast agent for tracking immortalized human bone marrow stromal cells in stroke treatment in rat. They used Effectene to transfect the MRI contrast agent Gd-DTPA into stem cells, because it is relatively more efficient and less toxic than liposome, calcium phosphate, or viral vectors for transfection into primary cells (188-190).

Many experiments have used other paramagnetic substances to track cell movement under MRI, including iron containing agents (e.g. Feridex) and a new generation of contrast agents magnetodendrimers and particles conjugated with Tat peptides (191). However, magnetodendrimers and particles conjugated with Tat peptides require complicated methods of manufacture. Furthermore, iron-containing agent labeled stem cells transplanted into brain are visualized as signal void images (black signal) using these paramagnetic substances. This can create problems because enlarged “false” black signaling effects of iron-labeled cells can exceed true stem cell mass (191). In addition, iron can be toxic at high concentrations (192), with its accumulation in tissue also catalyzing the Fenton reaction and potentiating oxygen toxicity through the
Figure 6. A typical MRI set from a representative animal showing angiogenesis. The temporal evolution of $K_i$ (blood-to-brain transfer constant) (a), CBF (b), CBV (c), $T_1$ (d), $T_{1\text{sat}}$ ($T_1$ in the presence of an off-resonance irradiation of the macromolecules of brain) (e), $T_2$ (f), and $K_{inv}$ (the inverse of the apparent forward transfer rate for magnetization transfer) (g) maps, respectively, obtained at various times from 1 day before to 6 weeks after cell transplantation. Adapted with permission from ref. 197.

generation of a wide range of free radical species. Furthermore, leakage of iron or the death of labeled cells may cause the release of iron oxide crystals into tissue, which can result in a potentially toxic uptake in surrounding healthy cells (193). In contrast, the pharmacological properties of Gd-DTPA have been extensively investigated (189) and clinically applied. Under MRI, Gd-DTPA–labeled cells in the brain show increased signal intensity (white signal) rather than void signals. Although the MR detection thresholds in stem cell labeling were lower in the iron-containing particle than that of Gd-DTPA it is highly desirable for clinical applications. In this study cortical neurochemical activity as evaluated by MRS also increased considerably after immortalized human bone marrow stromal cell transplantation.

Cell transplantation therapy for stroke holds great promise. However, many fundamental questions related to the optimal candidate (including the patient age, etiology, anatomic location and size of the infarct, and medical history), the best cell type, the number and concentration of cells, the timing of surgery, the route and site of delivery, and the need for immunosuppression remain to be answered. Furthermore, longer-term studies are required to determine whether the cell-enhanced recovery is sustained and also to determine the tumorigenic potential of the cells.
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Clearly, more research is needed to understand the bidirectional interaction between the transplanted cells and the host in which the role of molecular imaging is so important.

5. SUMMARY AND PERSPECTIVE

The development of therapies that will provide substantial beneficial outcome for stroke patients remains in its infancy. As of today, there is only one drug approved by FDA, recombinant tissue plasminogen activator, for the treatment of ischemic stroke. Thus, intense efforts to develop new therapeutics for the disease are inevitable. The great challenge for the development of newer, more effective acute stroke therapies will be to intertwine basic science and clinical trial methodology advances to maximize therapeutic advances for this increasingly common disorder. Translational research that focuses on taking advances in the basic science related to focal brain ischemia and developing therapeutic benefits from them will form the foundation for the future of acute stroke care.

The neuroprotection hypothesis, that agents interfering in the ischemic cascade of cell injury can protect the brain in acute stroke, has faced a setback by the latest news that NXY-059 failed to show benefit in the SAINT II trial (194). The main reasons for its failure was the fact that the site and mechanism of action of NXY-059 were never clearly delineated in an animal model (195, 196), so its mode of action in humans was never understood before moving into the clinic.

A variety of new agents have been shown most recently to improve post-ischemic outcomes in experimental animal models. During the preclinical stage, drug development research relies on animals to evaluate the efficacy of new experimental drugs before clinical trials. Typically since noninvasive preclinical molecular imaging of small animal models of stroke has not been established and is not widely available, experimental studies rely on traditional methods in which animals must be sacrificed for evaluation at each stage of drug development cycle. This leads to unavoidable intersubject variability so that a large number of animals must be sacrificed during the drug development and evaluation process. In vivo noninvasive imaging techniques can help reduce the number of animals in the preclinical evaluation of drugs since each animal can serve as its own control, and longitudinal studies are possible.

On the other hand, there is an opportunity to stimulate the brain’s own restorative capacity to improve neurological function after stroke. There are both cell-based and pharmaceutical approaches that amplify these restorative reprocesses and lead to improved neurological function in the experimental animal. The brain has the ability to create new neural cells in the adult similar to neurogenesis in the embryo; these neuroblasts express protective proteins and respond to homing signals emerging from the vasculature in the injured tissue. The development of these restorative treatments and the amplification of brain plasticity are also being assisted by the use of Molecular imaging of angiogenesis and the signaling pathways (Figure 6).

Molecular imaging has a pivotal role in identification of effective therapeutic agents not only by providing a better understanding of underlying mechanisms and signaling pathways in neuronal damage following stroke but it can also help us in noninvasive in vivo monitoring of the efficacy of these new therapeutic agents.

Additional advances in the time window for successful treatment and maximization of functional deficit reduction can be anticipated with the widespread application of penumbral imaging and the implementation of multimodality treatment paradigms. Molecular imaging techniques are becoming ever more a part of our decision-making and treatment strategies. Refinement of these techniques and the increasing use of multimodal imaging provide not only expected findings that are useful in clinical care of the patient but also provide unexpected findings that have expanded our current knowledge of stroke pathophysiology. The future for developing acute stroke therapies looks increasingly bright.

7. Reference

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computed tomography; MRI: magnetic resonance imaging; US: ultrasound; tPA: tissue plasminogen activator; DWI: Diffusion-weighted imaging; ADC: apparent diffusion coefficient; PWI: perfusion-weighted imaging; MRA: magnetic resonance angiography; DSA: digital subtraction angiography; CDUS: Carotid Duplex ultrasound; TCD: transcranial Doppler; CBF: cerebral blood flow; CBV: cerebral blood volume; TTP: time to peak; MTT: mean transit time; CMRO2: metabolic rate of oxygen consumption; OEF: rate of oxygen extraction; 18F-FMISO: 18F-fluoromisonidazole; MCAO: middle cerebral artery occlusion; NAA: \( \text{\textit{n}-acetyl aspartate; ppm: parts per million; AHA: American Heart Association; TE: echo time; MRS: MR spectroscopy; STEAM: stimulated echo acquisition mode; SNR: signal to noise ratio; AMPA: \( \alpha \)-amino-3-hydroxy-5-methyl-4-isoxazolone-propionic acid; NMDA: N-methyl-D-aspartate; SPIO: superparamagnetic iron oxide; MoAbs: monoclonal antibodies; USPIO: ultrassmall superparamagnetic iron oxide particles; PS: phosphatidylserine; DDC: N,N'-didansyl-L-cystine; HIF-1: hypoxia-inducible factor-1; VEGF: vascular endothelial growth factor; MSA: magnetic susceptibility artifacts; FEA: Finite element analysis; 1H-MRS: proton magnetic resonance spectroscopy.}

**Key Words:** Molecular Imaging, Stroke, Inflammation, Apoptosis, Angiogenesis, Stem cell therapy, Review

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